

Antimicrobial Susceptibility of Bacterial Pathogens Isolated from Canadian Intensive Care Units from 2007 to 2016: Results of the CANWARD Study

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Introduction

Subsequent to antimicrobial resistance data published from the Canadian National Intensive Care Unit (CAN-ICU) study (2005-2006) on 4,180 bacterial isolates from ICU patients (1), there has been little national surveillance data concerning antimicrobial susceptibility rates in Canadian ICUs. Antibiotic utilization and over-utilization, both in hospitals and the community, are important influences on the development of antibiotic-resistant pathogens such as MRSA, extended-spectrum β-lactamase-producing *Escherichia coli* and *Klebsiella* species, carbapenem-resistant *Enterobacteriaceae*, and multidrug-resistant *Pseudomonas aeruginosa* (1,2). It is estimated that 30-50% of antibiotic use in hospitals is unjustified (3). Each year in the United States, more than 2 million people become infected with bacteria that are resistant to antibiotics and at least 23,000 people die each year as a direct result of resistant infections (4). Infections caused by antibiotic-resistant organisms are associated with longer hospital stays, costly or prolonged treatments, and increased morbidity and mortality when compared to antibiotic-susceptible infections (5). It is estimated that 70% of ICU patients are on antibiotics at any one time (6).

The purpose of this prospective surveillance study was to evaluate the prevalence of infectious organisms, including resistant pathogens, and antimicrobial resistance patterns in Canadian ICUs from 2007 to 2016.

Materials and Methods

Bacterial Isolates: A total of 42,938 bacterial isolates (isolated from blood, urine, wound, and respiratory specimens) were submitted by tertiary-care medical centres from January 2007 to December 2016, inclusive, as part of the ongoing CANWARD national surveillance study. The medical centres were asked to submit clinical isolates (consecutive, one per patient, per infection site) that were clinically significant. Isolates were shipped to the coordinating laboratory (Health Sciences Centre, Winnipeg, Canada) where they were subcultured onto appropriate media and stocked in skim milk at -80° C. Of the isolates submitted, 8,130 (18.9%) were from patients admitted to an intensive care unit.

Antimicrobial Susceptibility Testing: Following 2 subcultures from frozen stock, *in vitro* antimicrobial susceptibility testing was performed using the Clinical and Laboratory Standards Institute (CLSI) broth microdilution method (7). Minimum inhibitory concentrations (MICs) were determined using custom, in-house prepared 96-well broth microdilution panels. Quality control was performed using CLSI recommended ATCC organisms. MIC interpretive criteria were defined according to CLSI breakpoints unless otherwise noted (8). Multidrug and extensive drug resistance (MDR, XDR) were defined as resistance to ≥3 or ≥5 antimicrobial agents, respectively.

Isolate Characterization: Potential MRSA isolates were confirmed by *mecA* PCR and further characterized by staphylococcal protein A (*spa*) typing to identify community-associated (CA-MRSA) and healthcare-associated (HA-MRSA) strains as previously described (9). Potential ESBL-producing isolates were confirmed by the CLSI confirmatory disc test. Potential vancomycin-resistant *Enterococcus* (VRE) isolates were confirmed by *van*/*vanB* PCR, as previously described (10). *Streptococcus pneumoniae* isolates were serotyped using commercial antisera. All statistical analysis was performed using the Cochran-Armitage test of trend.

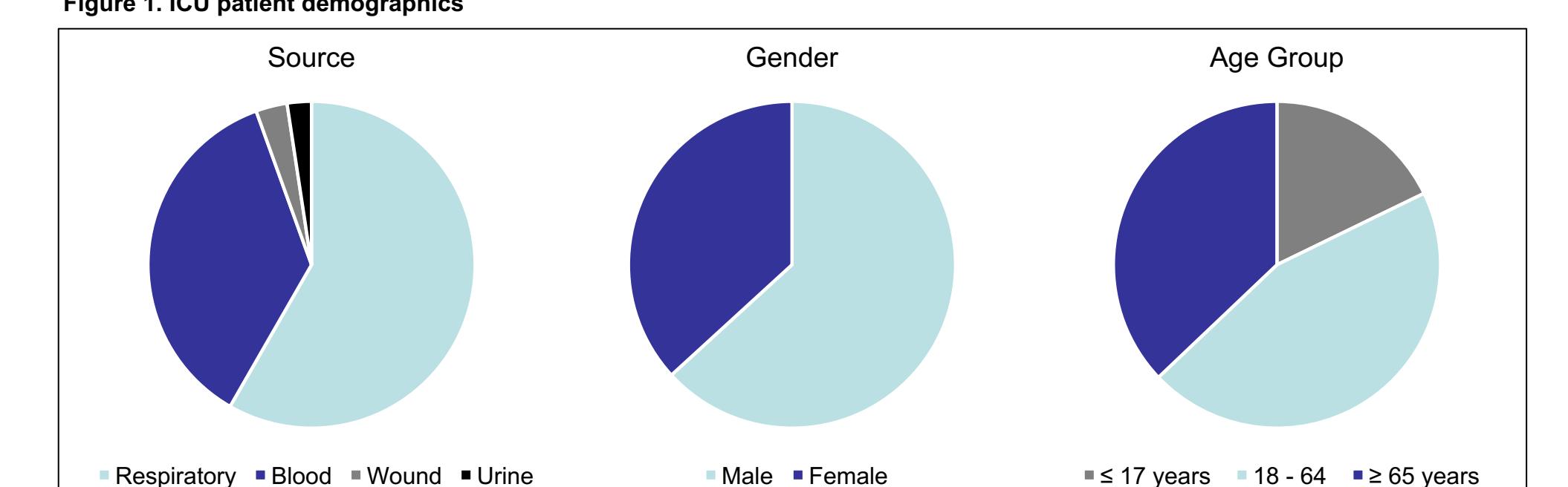
Results

Table 1. Antimicrobial susceptibility testing results of Gram-negative and Gram-positive pathogens collected from Canadian ICUs

Organism (n)	MIC (µg/ml)					Organism (n)	MIC (µg/ml)					Organism (n)	MIC (µg/ml)					Organism (n)	MIC (µg/ml)												
	Antimicrobial	%S	%I	%R	MIC ₅₀	MIC ₉₀	Min	Max	Antimicrobial	%S	%I	%R	MIC ₅₀	MIC ₉₀	Min	Max	Antimicrobial	%S	%I	%R	MIC ₅₀	MIC ₉₀	Min	Max							
<i>P. aeruginosa</i> (862)									<i>E. coli</i> (848)								<i>K. pneumoniae</i> (522)														
Amikacin	96.1	1.7	2.2	4	16	≤ 1	> 64	AMC	99.2	0.8	-	≤ 2	4	≤ 2	32	AMC	91.8	4.5	3.7	2	8	0.5	≥ 32	H. influenzae (425)							
Aztreonam	69.1	12.1	18.8	8	32	≤ 0.12	> 64	Cefazolin (IV)	60.5	12.2	27.3	2	> 128	0.5	> 128	Cefazolin	80.4	6.0	13.6	1	16	≤ 0.5	> 128	AMC	99.5	-	0.5	0.5	2	≤ 0.06	8
Cefepime	79.1	15.0	5.9	4	16	≤ 0.25	> 64	Cefotaxime	88.3	5.4	6.3	4	16	≤ 0.06	> 32	Cefotaxime	94.1	0.7	5.2	≤ 0.25	1	≤ 0.25	> 32	Ceftriaxone	97.2	2.3	0.5	1	2	≤ 0.25	> 16
Ceftazidime	75.3	7.9	16.8	4	32	≤ 0.25	> 32	Ceftazidime	87.9	1.7	10.3	≤ 0.5	16	≤ 0.25	> 32	Ceftazidime	93.7	-	6.3	≤ 0.25	1	≤ 0.25	> 64	Ciprofloxacin	93.7	1.9	4.4	≤ 0.06	0.5	≤ 0.06	> 16
Ceftobiprole	NB	-	4	16	0.5	128	CEFTOBIPROLE	85.4	0.7	13.9	≤ 0.25	32	≤ 0.25	> 256	Ceftriaxone	93.7	-	6.3	≤ 0.25	1	≤ 0.25	> 64	Ciprofloxacin	93.7	1.9	4.4	≤ 0.06	0.5	≤ 0.06	> 16	
C/T	97.5	1.3	0.5	2	≤ 0.12	> 64	Ciprofloxacin	72.0	0.4	27.7	≤ 0.06	> 16	≤ 0.06	> 16	Ceftriaxone	93.7	-	6.3	≤ 0.25	1	≤ 0.25	> 64	Ciprofloxacin	93.7	1.9	4.4	≤ 0.06	0.5	≤ 0.06	> 16	
Ciprofloxacin	76.0	8.0	16.0	0.25	4	≤ 0.06	> 16	Ciprofloxacin	72.0	0.4	27.7	≤ 0.06	> 16	≤ 0.06	> 16	Ceftriaxone	93.7	-	6.3	≤ 0.25	1	≤ 0.25	> 64	Ciprofloxacin	93.7	1.9	4.4	≤ 0.06	0.5	≤ 0.06	> 16
Colistin	94.7	-	5.3	1	2	≤ 0.06	> 16	Colistin	99.7	-	0.3	0.25	0.5	≤ 0.06	8	Colistin	97.7	-	2.3	0.5	1	≤ 0.06	> 16	Daptomycin	99.5	-	0.5	0.5	1	≤ 0.25	4
Gentamicin	83.9	6.7	9.4	2	8	≤ 0.5	> 32	Ertapenem	99.0	0.4	0.7	0.03	0.06	≤ 0.03	> 32	Ertapenem	99.0	0.6	0.4	≤ 0.03	0.06	≤ 0.03	> 32	Ertapenem	99.5	-	0.5	0.3	0.12	≤ 0.03	> 4
Imipenem	62.9	8.7	28.5	2	32	≤ 0.03	> 32	Gentamicin	87.5	0.7	11.8	≤ 0.5	32	≤ 0.5	> 32	Gentamicin	96.7	-	3.3	≤ 0.5	0.5	≤ 0.5	> 32	Gentamicin	96.7	-	3.3	≤ 0.5	0.5	≤ 0.5	> 2
Meropenem	73.7	8.0	18.3	1	16	≤ 0.03	> 64	Meropenem	99.9	-	0.1	0.03	0.12	≤ 0.03	32	Meropenem	99.6	0.2	0.2	≤ 0.03	0.06	≤ 0.03	> 16	Meropenem	99.7	-	0.3	0.06	0.12	≤ 0.06	2
Tobramycin	93.3	0.7	6.0	≤ 0.5	2	≤ 0.5	> 64	Moxifloxacin	NB	-	0.06	≤ 16	≤ 0.06	≤ 16	Moxifloxacin	95.0	2.1	2.9	2	8	≤ 1	> 512	TZP	95.0	2.1	2.9	2	8	≤ 1	> 512	
TZP	75.8	14.4	9.9	8	64	≤ 1	> 512	Tigecycline	94.2	1.9	3.9	2	8	≤ 1	> 512	Tigecycline	93.7	5.0	1.3	2	2.5	≤ 0.12	> 8	Tigecycline	93.7	5.0	1.3	2	2.5	≤ 0.12	> 8
SXT	70.5	-	29.5	≤ 0.12	> 8	≤ 0.12	> 8	SXT	92.7	-	7.3	≤ 0.12	1	≤ 0.12	> 8	SXT	82.6	4.0	13.4	≤ 0.12	4	≤ 0.12	> 8	SXT	82.6	4.0	13.4	≤ 0.12	4	≤ 0.12	> 8

^a, percent susceptibility interpreted using EUCAST breakpoints. ^b, percent susceptibility interpreted using FDA breakpoints. MSSA: methicillin-susceptible *S. aureus*, MRSA: methicillin-resistant *S. aureus*, AMC: amoxicillin/clavulanate, C/T: ceftolozane/tazobactam, TZP: piperacillin/tazobactam, SXT: trimethoprim/sulfamethoxazole, NB: no breakpoints available.

Figure 1. ICU patient demographics



This study identified MDR/XDR in the following: 19.0/2.3% of *S. aureus*, 5.1/1.9% of *S. pneumoniae*, 15.0/2.0% of *P. aeruginosa*, 26.3/14.9% of *E. coli* and 8.6/5.4% of *K. pneumoniae*. The proportion of MDR/XDR *E. coli* increased significantly over the study period, from 13.4/3.2% in 2007 to 30.4/18.8% in 2016 ($P<0.0001$). MDR/XDR *K. pneumoniae* also increased significantly in prevalence, from 2.5/0% in 2007 to 13.6/12.8% in 2016 ($P=0.022$). The most common *S. pneumoniae* serotypes isolated from Canadian ICUs were 3 (9.1%) and 11A (8.0%), predominantly associated with respiratory infections.

Rank	Organism	N	% Total
1	<i>Staphylococcus aureus</i>	1746	21.5
2	<i>Pseudomonas aeruginosa</i>		